

Research Article

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Lactic Acid Bacteria from Tarkhineh, A Traditional Iranian Fermented Cereal Product Investigated by Cultivation and 16S rRNA Clone Library. A Short Communication.

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Abstract

The present study address to characterize the bacterial community of Tarkhineh, a traditional Iranian fermented cereal product, by conventional culturing and 16S rRNA clone library. Twenty-nine lactic acid bacteria (LAB) from culturing on Man, Rogosa and Sharpe agar were identified by 16S rRNA gene sequencing and 55 clones from the 16S rRNA clone library. Discrepancies regarding the identity of the LAB were encountered by the two methods. By cultivation, Lactobacillus (Lb.) plantarum and Lb. casei were dominant followed by Lb. brevis, Lb. paracasei, Lb. paracasei subsp. paracasei, Lb. pentosus, Lb. zeae and uncultured bacterium clone MS-238. In contrast, construction of the clone library analysis revealed that the dominant clones belonged to Lb. paralimentarius, but Lb. alimentarius, Lb. delbrueckii subsp. bulgaricus, Lb. kefiri, Lb. kefiranofaciens subsp. kefirgranum, Lb. pontis, Lactobacillus sp. and Pediococcus were also detected. In addition to LAB, seven clones belonging to Bacillus and three bacterium clones were also detected.

Keywords: Tarkhineh, Lactic Acid Bacteria, Cultivation and 16S rRNA Clone Library.

Introduction

According to Ozen and Dinleyici, sophisticated technology in molecular archeology, can trace the use of fermented products as early as 8.000 BC [1]. Tarkhineh is the basic part of a traditional Iranian fermented cereal soup in the west mountain area of Iran. It is compost of grind wheat, soak in fermented milk and the local people use this soup in winter to prevent influenza and reduce symptoms. Based on processing of Tarkhineh it seems that the product could be a good source of lactic acid bacteria (LAB) with remedy functions. Some previous studies have investigated the microbial communities in Tarkhineh by cultivation, but culturedependent methods do not reflect the true bacteriological community, rather the needs of different growth media for cultivation of certain bacteria [2-6]. Therefore, various culture-independent methods are developed during the last decades e.g., including 16S rRNA clone libraries [7, 8]. Knowledge is accessible on the construction of 16S rRNA clone libraries to evaluate the bacterial diversity of fermented food and milk products e.g., [9-13]. The aim of the present study was construction of a 16S rRNA clone library and compare the results with 16S rRNA sequence analysis of culturable bacteria from Tarkhineh.

Materials and Methods Isolation of Bacterial Strains

Twenty grams of Tarkhineh were inoculated into 200 mL of man, Rogosa and Sharpe (MRS) broth (Fuka, catalogue no. 69966). Isolation of bacteria in tarkhineh was carried out according to the procedure described elsewhere [14]. To avoid time consuming isolation steps and interference with yeast, lactobacilli colonies were selected on MRS agar containing nystatin as suggested elsewhere [15]. Plates were incubated under anaerobic conditions at 37°C for 48 to 72 h and Gram-positive rods and catalase negative were cultured in MRS broth and stored at -80oC until further use.

Purification of bacterial DNA and 16S rRNA gene sequence analysis of culturable bacteria DNA was extracted from 29 randomly selected isolates by a method described elsewhere [16]. The pellet was resuspended in 100 μ L deionized water and the concentration of DNA was determined spectrophotometrically at wavelength of 260nm. The DNA was stored at -20°C until further use. The selected strains were identified by 16S rRNA gene sequence analysis as described elsewhere [17].

Construction of 16S rRNA Clone Library

Bacterial DNA was purified with QIAmp DNA Minikit (Qiagen), according to the manufactures protocol for bacterial DNA. The clone library was constructed as previously described [12, 13]. Of the 98 clones isolated, 69 had an insert and amplified 16S rRNA genes of 55 clones were sequenced. Sequences were individually analyzed and edited in the program Chromas Pro. Edited sequences were subjected to Blast search in the NCBI database and subsequently all samples were aligned with selected reference sequences in the BioEdit program.

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Results

16S rRNA gene sequencing of culturable bacteria

The total viable count of the Tarkhineh sample was between $4.4 \ge 10^5 - 6.8 \ge 10^5$ CFU g⁻¹. Twenty-nine LAB strains were randomly selected and examined by microscopy after Gram staining, catalase and oxidase reaction and further characterized by 16S rRNA gene sequencing. The LAB strains and their corresponding accession no. revealed that the culturable bacteria were dominated by strains showing high similarities to Lactobacillus (Lb.) plantarum (11 isolates), Lb. casei (six isolates), Lb. brevis (four isolates), Lb. paracasei (two isolates), one isolate to Lb. pentosus, Lb. zeae (one isolate) and one isolate showing similarity to uncultured bacterium clone MS-238 (Table 1).

Table 1: Identification of Culturable Bacteria Isolated from Tarkhineh with Partial Sequence of 16S rRNA GenesReferenced to Accession no. in GenBank

| Strain no. | Closest relative (obtained from Blast search) | Accession no. | Similarity (%) |
|---------------|---|---------------|----------------|
| T1 | Lactobacillus plantarum isolate L4-1 | AB550299 | 98 |
| Т2 | Lactobacillus plantarum strain S2 | GU292429 | 95 |
| Т3 | Lactobacillus plantarum strain KLDS 1.0725 | EU626010 | 97 |
| T4 | Lactobacillus plantarum isolate L4-1 | AB550299 | 98 |
| Т6 | Lactobacillus plantarum | AB494717 | 97 |
| Т8 | Lactobacillus plantarum strain IMAU:10272 | GU138600 | 99 |
| T10 | Lactobacillus plantarum KLDS 1.0725 | EU626010 | 91 |
| T16 | Lactobacillus plantarum strain IMAU700004 | GQ131121 | 98 |
| T19 | Lactobacillus plantarum | AB510751 | 98 |
| T23 | Lactobacillus plantarum | AB494717 | 90 |
| T26 | Lactobacillus plantarum | AB494717 | 98 |
| Т9 | Lactobacillus casei | AB494735 | 95 |
| T13 | Lactobacillus casei | AB494735 | 93 |
| T17 | Lactobacillus casei | AB494735 | 97 |
| T18 | Lactobacillus casei | AB494735 | 97 |
| T20 | Lactobacillus casei | AB494735 | 97 |
| T25 | Lactobacillus casei | AB494735 | 91 |
| T11 | Lactobacillus brevis | GU369768 | 91 |
| T14 | Lactobacillus brevis | GU138534 | 98 |
| T15 | Lactobacillus brevis strain JS-7-3 | GU369768 | 94 |
| T24 | Lactobacillus brevis | AB494718 | 96 |
| Т5 | Lactobacillus paracasei subsp. paracasei | AY773951 | 94 |
| T12 | Lactobacillus paracasei subsp. paracasei | FJ861109 | 94 |
| T29 | Lactobacillus paracasei subsp. paracasei tot. 3 | AY773951 | 96 |
| T21 | Lactobacillus paracasei strainKLDS1.0658 | FJ607302 | 96 |
| T28 | Lactobacillus paracasei strainKLDS1.0658 | FJ607302 | 95 |
| Τ7 | Lactobacillus pentosus strain N3 | GU253891 | 99 |
| T27 | Lactobacillus zeae | AB362765 | 98 |
| T22 | Uncultured bacterium clone MS-238 | GQ477897 | 99 |

Volume

16S rRNA clone library

Fifty-five clones from the clone library were sequenced and revealed that clones belonging to LAB were dominant and they belonged to: Lb. paralimentarius, 31 clones, one clone to Lb. alimentarius, three clones to Lb. delbrueckii subsp. bulgaricus, Lb. kefiri (two clones), Lb. kefiranofaciens subsp. kefirgranum (one clone), Lb. pontis (one clone), six clones to Lactobacillus and one clone to Pediococcus parvulus. In addition, were six cones belonged to Bacillus and three uncultured clones detected (Table 2).

Table 2: Identification of Clones from Tarkhineh with Partial Sequence of 16S rRNA Genes Referenced to Accession no. in GenBank

| Closest relative (obtained from Blast search) | Accession no. | Similarity (%) | No. of clones showing high similarity to the closest relative |
|---|---------------|----------------|---|
| Lactobacillus paralimentarius strain 412 | EU483111 | 98 | 30 |
| Lactobacillus paralimentarius strain DSM 13238 | AJ417500 | 97 | 1 |
| Lactobacillus alimentarius strain M-M3 | FJ4157241 | 97 | 1 |
| Lactobacillus delbrueckii subsp. bulgaricus ATCC BAA- 365 | CP000412 | 96 | 1 |
| Lactobacillus delbrueckii subsp. bulgaricus strain IMAU20102 | FJ845002 | 96 | 1 |
| Lactobacillus delbrueckii subsp. bulgaricus strain IMAU40079 | FJ749354 | 96 | 1 |
| Lactobacillus kefiri strain IMAU450007 | FJ749410 | 96 | 1 |
| Lactobacillus kefiri strain YIT 0222 | AB429371 | 97 | 1 |
| Lactobacillus kefiranofaciens subsp. kefirgranum strain IMAU50004 | FJ749408 | 96 | 1 |
| Lactobacillus pontis strain LMG 114187 | AJ422032 | 98 | 1 |
| Lactobacillus genome sp. C2 | AY278620 | 98 | 1 |
| Lactobacillus sp. | | <96 | 5 |
| Pediococcus parvulus strain LA1110 | FM878597 | 98 | 1 |
| Bacillus cereus Q1 | CP000227 | 96 | 1 |
| Bacillus subtilis strain E9-1 | FJ573171 | 97 | 1 |
| Bacillus subtilis strain RC24 | FJ263368 | 96 | 1 |
| Uncultured Bacillus sp. clone W2 | FJ863099 | 97 | 1 |
| Bacillus sp. | | <96 | 1 |
| Brevibacillus thermoruber | AB362290 | 96 | 1 |
| Uncultured bacterium clone p-2370-55G5 | AF371474 | 97 | 1 |
| Uncultured bacterium clone p-3083-SwA-3 | AF371484 | 97 | 1 |
| Uncultured bacterium clone AP07K.42 | AM275437 | 97 | 1 |

Discussion

Tarkhineh fermentation occurs mainly by bacteria naturally present in the raw materials included flour, dough, salt, turnip, leaven (mint, red pepper) containing numerous microorganisms including LAB [3, 4]. Due to the popularity of Tarkhineh and its increasing consumption some information is known about the bacterial community of the product [2-6]. In these studies, the bacterial community was evaluated by traditional cultivation. However, as culture-dependent methods provide a rather skewed and restricted presentation of the bacteriological community, we investigated the bacterial community in Tarkhineh by cultivation and 16S rRNA clone library. Of the culturable LAB strains isolated in the present study, were Lb. brevis, Lb. casei, Lb. paracasei, Lb. pentosus and Lb. zeae, of which culturable Lb. brevis, Lb. casei and Lb. pentosus strains previously reported in Tarkhineh [5, 18, 19]. In addition, one of the culturable bacteria isolated in the current study, uncultured bacterium clone MS-238, a strain previously isolated from olive-oil mill wastewater revealing high similarity to Lb. rhamnosus [20].

Clone library analysis of Tarkhineh revealed several LAB strains, Lb. alimentarius, Lb. delbrueckii, Lb. kefiri, Lb. kefiranofaciens, Lb. paralimentarius and Lb. pontis not frequently isolated from Iranian fermented products. The discrepancy determined in the bacterial community determined by culture-dependent and independent methods of Tarkhineh in the present study is a finding previously been reported in variety ecosystems and fermented milk, koumiss from Mongolia [13, 21-25]. 16S rRNA clone library analysis might be more representative of the community in qualitative and quantitative terms especially when the

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numbers of clones were enough. Even though relative few bacteria strains were isolated, and few clones investigated, combining partial sequencing of 16S rRNA genes of the culturable bacteria and the 16S rRNA clone library, interesting results were obtained showing a diverse LAB community of the investigated Tarkhineh sample.

Interesting findings reported in previous studies revealing inhibitory effect of LAB isolated from Tarkhineh towards Escherichia coli and Listeria monocytogenes and E. coli, Pseudomonas aeruginosa. Salmonella typhimurium and Staphylococcus aureus, which S. aureus was the most susceptible strain [6, 26]. These studies and some other studies revealing probiotic potential characteristics of LAB indicating that LAB isolated from Tarkhineh can be used as suitable supplements to functional foods [19, 27-29]. In addition to the presence of potential probiotics reported associated to Tarkhineh it is of interest noticing that fermentation of Tarkhineh by Lb. bulgaricus and Streptococcus thermophilus as well as the native probiotics in Tarkhineh binds mycotoxins and thereby reducing the concentrations of aflatoxin M1 and B1 during fermentation [30].

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References

- 1. Ozen, M., Dinleyici, E. C. (2015). The history of probiotics: the untold story. *Beneficial microbes*, *6*(2), 159-165.
- 2. Ghaitaranpour, A., Yazdi, F. T., Behbahani, B. A., Mortazavi, A., Mohebbi, M. (2013). The effect of wheat boiling time, bulgur particle size, drying time and temperature on some physical properties of hot air dried Tarkhineh. *Scientific Journal of Pure and Applied Sciences, 2*(4), 175-182.
- 3. Tabatabaei Yazdi, F., Alizadeh Behbahani, B., Mohebbi, M., Mortazavi, S. A., Ghaitaranpour, A. (2012). Identification of lactic acid bacteria isolated from Tarkhineh, a traditional Iranian fermented food. *Scientific Journal of Microbiology*, 1(7), 152-159.
- Tabatabaei-Yazdi, F., Alizadeh-Behbahani, B., Mortazavi, A. (2013). Effect of temperature and salt concentration on microbial changes during tarkhineh fermentation Scientific Journal of Biological Science, 2, 8-16.
- 5. Vasiee, A. R., Mortazavi, A., Tabatabaei-yazdi, F., Dovom, M. R. (2018). Detection, identification and phylogenetic analysis of lactic acid bacteria isolated from Tarkhineh, Iranian fermented cereal product, by amplifying the 16s rRNA gene with universal primers and differentiation using rep-PCR. *International Food Research Journal*, *25*(1).
- Vasiee, A., Falah, F., Behbahani, B. A., Tabatabaee-Yazdi, F. (2020). Probiotic characterization of Pediococcus strains isolated from Iranian cereal-dairy fermented product: Interaction with pathogenic bacteria and the enteric cell line Caco-2. *Journal of Bioscience and Bioengineering,*

130(5), 471-479.

- 7. Theron, J., Cloete, T. E. (2000). Molecular techniques for determining microbial diversity and community structure in natural environments. *Critical Reviews in Microbiology*, *26*(1), 37-57.
- 8. Jany, J. L., Barbier, G. (2008). Culture-independent methods for identifying microbial communities in cheese. *Food Microbiology*, *25*(7), 839-848.
- 9. Callon, C., Duthoit, F., Delbès, C., Ferrand, M., Le Frileux, Y., et al. (2007). Stability of microbial communities in goat milk during a lactation year: molecular approaches. *Systematic and Applied Microbiology*, *30*(7), 547-560.
- Rasolofo, E. A., St-Gelais, D., LaPointe, G., Roy, D. (2010). Molecular analysis of bacterial population structure and dynamics during cold storage of untreated and treated milk. *International Journal of Food Microbiology*, *138*(1-2), 108-118.
- 11. Raats, D., Offek, M., Minz, D., Halpern, M. (2011). Molecular analysis of bacterial communities in raw cow milk and the impact of refrigeration on its structure and dynamics. *Food Microbiology*, *28*(3), 465-471.
- Osvik, R. D., Sperstad, S., Breines, E. M., Godfroid, J., Zhou, Z., et al. (2013). Bacterial diversity of amasi, a South African fermented milk product, determined by clone library and denaturing gradient gel electrophoresis analysis African Journal of Microbiology Research, 7(32), 4146-4158.
- Ringø, E., Andersen, R., Sperstad, S., Zhou, Z., Ren, P., et al. (2014). Bacterial community of koumiss from Mongolia investigated by culture and culture-independent methods. *Food Biotechnology*, 28(4), 333-353.
- 14. Pennacchia, C., Ercolini, D., Blaiotta, G., Pepe, O., Mauriello, G., et al. (2004). Selection of Lactobacillus strains from fermented sausages for their potential use as probiotics. *Meat Science*, *67*(2), 309-317.
- Gardiner, G. E., Casey, P. G., Casey, G., Lynch, P. B., Lawlor, P. G., et al. (2004). Relative ability of orally administered Lactobacillus murinus to predominate and persist in the porcine gastrointestinal tract. *Applied and Environmental Microbiology*, 70(4), 1895-1906.
- 16. Pitcher, D. G., Saunders, N. A., Owen, R. J. (1989). Rapid extraction of bacterial genomic DNA with guanidium thiocyanate. *Letters in Applied Microbiology*, *8*(4), 151-156.
- Ringø, E., Sperstad, S., Myklebust, R., Refstie, S., Krogdahl, Å. (2006). Characterisation of the microbiota associated with intestine of Atlantic cod (Gadus morhua L.): the effect of fish meal, standard soybean meal and a bioprocessed soybean meal. *Aquaculture*, 261(3), 829-841.
- Tafvizi, F., Tajabadi Ebrahimi, M. (2015). Application of repetitive extragenic palindromic elements based on PCR in detection of genetic relationship of lactic acid bacteria species isolated from traditional fermented food products. *Journal of Agricultural Science and Technology*, 17(1), 87-98.
- 19. Falah, F., Vasiee, A., Yazdi, F. T., Behbahani, B. A. (2021). Preparation and functional properties of synbiotic yogurt fermented with Lactobacillus brevis pml1 derived from a fermented cereal-dairy product. *BioMed*

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research international, 2021(1), 1057531.

- Kavroulakis, N., Ntougias, S. (2011). Bacterial and β-proteobacterial diversity in Olea europaea var. mastoidis-and O. europaea var. koroneiki-generated olive mill wastewaters: influence of cultivation and harvesting practice on bacterial community structure. *World Journal of Microbiology and Biotechnology*, 27, 57-66.
- Tamaki, H., Sekiguchi, Y., Hanada, S., Nakamura, K., Nomura, N., et al. (2005). Comparative analysis of bacterial diversity in freshwater sediment of a shallow eutrophic lake by molecular and improved cultivationbased techniques. *Applied and Environmental Microbiology*, 71(4), 2162-2169.
- 22. Michaud, L., Lo Giudice, A., Troussellier, M., Smedile, F., Bruni, V., et al. (2009). Phylogenetic characterization of the heterotrophic bacterial communities inhabiting a marine recirculating aquaculture system. *Journal of Applied Microbiology*, 107(6), 1935-1946.
- 23. Huang, Z. B., Guo, F., Zhao, J., Li, W. D., Ke, C. H. (2010). Molecular analysis of the intestinal bacterial flora in cage-cultured adult small abalone, Haliotis diversicolor. *Aquaculture Research*, *41*(11), e760-e769.
- 24. Zeng, Y., Ma, Y., Wei, C., Jiao, N., Tang, K., et al. (2010). Bacterial diversity in various coastal mariculture ponds in Southeast China and in diseased eels as revealed by culture and culture-independent molecular techniques. *Aquaculture Research*, 41(9), e172-e186.
- 25. Szabó, G., Khayer, B., Rusznyák, A., Tátrai, I., Dévai, G., et al. (2011). Seasonal and spatial variability of sediment

bacterial communities inhabiting the large shallow Lake Balaton. *Hydrobiologia, 663,* 217-232.

- Ebrahimi, M. T., Shariatpanahi, M., Jafari, P., Sadeghi, S. (2014). Inhibitory effect of Lactobacilli (isolated from Tarkhineh Dough) on E. coli and Lis. monocytogenes colonization International Journal of Biosciences, 5(10), 29-36.
- Kiani, A., Nami, Y., Hedayati, S., Elieh Ali Komi, D., Goudarzi, F., et al. (2021). Application of Tarkhineh fermented product to produce potato chips with strong probiotic properties, high shelf-life, and desirable sensory characteristics. *Frontiers in Microbiology*, *12*, 657579.
- Kiani, A., Nami, Y., Hedayati, S., Jaymand, M., Samadian, H., et al. (2021). Tarkhineh as a new microencapsulation matrix improves the quality and sensory characteristics of probiotic Lactococcus lactis KUMS-T18 enriched potato chips. *Scientific Reports, 11*(1), 12599.
- Salek, F., Mirzaei, H., Khandaghi, J., Javadi, A., Nami, Y. (2023). Apoptosis induction in cancer cell lines and anti-inflammatory and anti-pathogenic properties of proteinaceous metabolites secreted from potential probiotic Enterococcus faecalis KUMS-T48. *Scientific Reports*, *13*(1), 7813.
- 30. Moradi, L., Paimard, G., Sadeghi, E., Rouhi, M., Mohammadi, R., et al. (2022). Fate of aflatoxins M1 and B1 within the period of production and storage of Tarkhineh: A traditional Persian fermented food. *Food Science Nutrition*, 10(3), 945-952.